

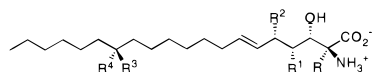
A New Strategy for the Synthesis of Sphingosine Analogues. Sphingofungin F

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Sphingosines, compounds consisting of polar polyhydroxyl amino head groups and long lipid chains, are membrane constituents involved in a number of cellular events including protein binding (GPI anchors) and transmembrane signaling.¹ A related series of compounds wherein the primary alcohol is oxidized to a carboxylic acid such as sphingofungin B (**1**)² or possesses a



- 1 R=H, R¹=R²=R³=OH, R⁴=H
- 2 R=CH₃, R¹=R²=OH, R³=R⁴=O
- 3 R=CH₂OH, R¹=OH, R²=H, R³=R⁴=O
- 4 R=CH₂OH, R¹=R²=H, R³=R⁴=O

quaternary center such as sphingofungin F (**2**)³ were found to inhibit the biosynthesis of sphingolipids due to their activity as serinepalmitoyl transferase inhibitors.⁴ These compounds are also strikingly similar to myriocin (**3**),⁵ a compound shown to be 10–100 times more potent than cyclosporin A.⁶ Mycestericin D (**4**), a deoxy analogue, and its dihydro and 3-*epi* isomer have also been isolated.⁷ The biological importance of these compounds stimulated a number of synthetic efforts largely making use of the “chiral pool”.⁸ The difficulties of creating quaternary centers asymmetrically in catalytic procedures and the noted biological activity of these analogues led us to develop a general strategy to this series. We now report the successful realization of a

(1) Reviews: (a) Hannun, Y. A. *Sphingolipid-Mediated Signal Transduction*; Chapman & Hall: New York, NY, 1997. (b) Merrill, A. H., Jr.; Sweeley, C. C. In *Biochemistry of Lipids, Lipoproteins and Membranes*; Vance, D. E., Vance, J., Eds.; Elsevier Science B. V.: Amsterdam, 1996; pp 309–339. (c) Hakomori, S. *Sphingolipid Biochemistry*. In *Handbook of Lipid Research*; Kafner, J. N., Hakomori, S., Eds.; Plenum: New York, 1983; Vol. 3, p 1. (d) Ariga, T.; Jarvis, W. D.; Yu, R. K. *J. Lipid Res.* **1998**, *39*, 1. (e) Igarashi, Y. *J. Biochem.* **1997**, *122*, 1080. (f) Hannun, Y. A. *Science* **1996**, *274*, 1855. (g) Shayman, J. A. *J. Am. Soc. Nephrol.* **1996**, *7*, 171. (h) Spiegel, S.; Milstein, S. *J. Membr. Biol.* **1995**, *146*, 225. See, also: (i) Nugent, T. C.; Hudlicky, T. *J. Org. Chem.* **1998**, *63*, 510 and references cited therein. (j) Kobayashi, S.; Furuta, T.; Hayashi, T.; Nishijima, M.; Hanada, K. *J. Am. Chem. Soc.* **1998**, *120*, 908 and references cited therein.

(2) (a) VanMiddlesworth, F.; Giacobbe, R. A.; Lopez, M.; Garrity, G.; Bland, J. A.; Bartizal, K.; Fromtling, R. A.; Polishook, J.; Zweierink, M.; Edison, A. M.; Rozdilsky, W.; Wilson, K. E.; Monaghan, R. L. *J. Antibiot.* **1992**, *45*, 861. Structure elucidation: (b) VanMiddlesworth, F.; Dufresne, C.; Wincott, F. E.; Mosley, R. T.; Wilson, K. E. *Tetrahedron Lett.* **1992**, *33*, 297.

(3) Horn, W. S.; Smith, T. L.; Bills, G. F.; Raghoobar, S. L.; Helms, G. L.; Kurtz, M. B.; Marrinan, J. A.; Frommer, B. R.; Thromton, R. A.; Mandala, S. M. *J. Antibiot.* **1992**, *45*, 1692.

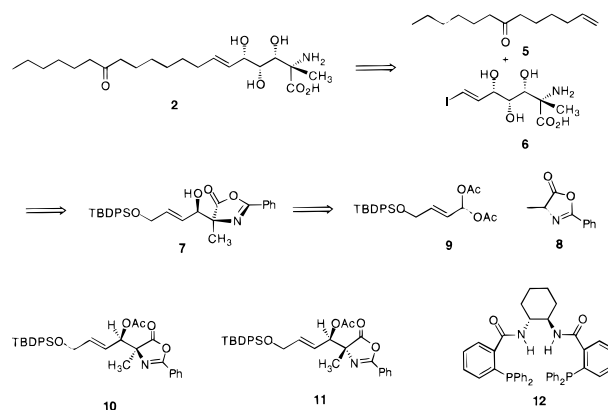
(4) Zweierink, M. M.; Edison, A. M.; Well, G. B.; Pinto, W.; Lester, R. L. *J. Biol. Chem.* **1992**, *267*, 25032.

(5) Myriocin (ISP-I, thermozymocidin) has been isolated from various strains, see: (a) Kluepfel, D.; Bagli, J.; Chrest, A. K.; Sehgal, C. V. *J. Antibiot.* **1972**, *25*, 109. Bagli, J. F.; Kluepfel, D. *J. Org. Chem.* **1973**, *38*, 1253. (b) Aragozzini, F.; Marachini, P. L.; Craveri, R. *Tetrahedron* **1972**, *28*, 5493. (c) Sasek, V.; Sailer, M.; Vokoun, J.; Musilek, V. *J. Basic Microbiol.* **1989**, *29*, 383. (d) Fujita, T.; Inoue, K.; Yamamoto, S.; Ikumoto, T.; Sasaki, S.; Toyama, R.; Yoneta, M.; Hoshino, Y.; Okumoto, T. *J. Antibiot.* **1994**, *47*, 208.

(6) (a) Reference 5d. (b) Miyake, Y.; Kozutsumi, Y.; Nakamura, S.; Fujita, T.; Kawasaki, T. *Biochem. Biophys. Res. Commun.* **1995**, *211*, 396.

(7) Fujita, T.; Hamamichi, N.; Kiuch, M.; Matsuzaki, T.; Kitao, Y.; Inoue, K.; Hirose, R.; Yoneta, M.; Sasaki, S.; Chiba, K.; Fujita, T.; Inoue, K.; Yamamoto, S.; Ikumoto, T.; Sasaki, S.; Toyama, R.; Yoneta, M.; Hoshino, Y.; Okumoto, T. *J. Antibiot.* **1996**, *49*, 846. Sasaki, S.; Hashimoto, R.; Yoneta, M.; Sasaki, S.; Inoue, K.; Ikumoto, T.; Hirose, R.; Chiba, K.; Hoshino, Y.; Okumoto, T.; Fujita, T. *J. Antibiot.* **1994**, *47*, 420.

Scheme 1. Retrosynthetic Analysis



concise synthesis of sphingofungin F in which all stereochemistry emanates from a new asymmetric alkylation in which an asymmetric palladium complex differentiates between enantiotopic leaving groups of a *gem*-diacetate and enantiotopic faces of an azlactone enolate.

Scheme 1 illustrates the retrosynthetic analysis whereby the major disconnection splits the molecule into the lipid tail **5** and the polar head **6**, the latter being the challenging fragment. If two of the hydroxyl groups derive from a distereoselective *cis*-dihydroxylation, the serine analogue **7** becomes a precursor. The stereochemistry of the C-3 hydroxyl group was inverted from the natural product to address the known effects of such allylic functionality on the diastereoselectivity of the osmium catalyzed dihydroxylation.⁹ Such quaternary serine analogues **7** may derive from our newly developed asymmetric alkylation of azlactones with *gem*-diacetates, in this case requiring **8** and **9**, respectively.^{10,11}

The *gem*-diacetate **9**, derived from the corresponding known aldehyde,¹² is available in two steps from commercially available *cis*-2-butene-1,4-diol, in quantitative yield by ferric chloride (0.1 mol%) catalyzed addition of acetic anhydride.¹¹ The Pd catalyzed alkylation must control both relative and absolute stereochemistry. For example, by using triphenylphosphine as the ligand, the two diastereomers **10** and **11** (as their racemates) are formed in a 1:1.6 ratio at room temperature. Thus, the chiral ligand must override the intrinsic bias of the substrates to provide **7**.

Performing the alkylation of the sodium salt of the azlactone **8** (NaH, THF) and *gem*-diacetate **9** with the catalyst derived from π -allylpalladium chloride dimer (0.5%) and ligand **12**¹³ (1.5%)

(8) For total synthesis of myriocin, see: (a) Banfi, L.; Bretta, M. G.; Colombo, L.; Gennari, C.; Scolastico, C. *J. Chem. Soc., Chem. Commun.* **1982**, 488. Banfi, L.; Bretta, M. G.; Colombo, L.; Gennari, C.; Scolastico, C. *J. Chem. Soc., Perkin Trans. 1* **1983**, 1613. (b) Yoshikawa, M.; Yokokawa, Y.; Okuno, Y.; Murakami, N. *Chem. Pharm. Bull.* **1994**, *42*, 994. Yoshikawa, M.; Yokokawa, Y.; Okuno, Y.; Murakami, N. *Tetrahedron* **1995**, *51*, 6209. (c) Sano, S.; Kobayashi, Y.; Kondo, T.; Takebayashi, M.; Maruyama, S.; Fujita, T.; Nagao, Y. *Tetrahedron Lett.* **1995**, *36*, 2097. (d) Hatakeyama, S.; Yoshida, M.; Esumi, T.; Iwabuchi, Y.; Irie, H.; Kawamoto, T.; Yamada, H.; Nishizawa, M. *Tetrahedron Lett.* **1997**, *45*, 7887. For formal synthesis of myriocin see: (e) Rao, A. V. R.; Gurjar, M. K.; Devi, T. R.; Kumar, K. R. *Tetrahedron Lett.* **1993**, *34*, 1653. (f) Deloisy, S.; Thang, T. T.; Olesker, A.; Lukas, G. *Tetrahedron Lett.* **1994**, *35*, 4783. For total synthesis of sphingofungin B and F, see: (g) Kobayashi, S.; Matsumura, M.; Furuta, T.; Hayashi, T.; Iwamoto, S. *Synlett* **1997**, 301 and ref 1j. For total synthesis of sphingofungin D, see: (h) Chida, N.; Ikemoto, H.; Noguchi, A.; Amano, S.; Ogawa, S. *Nat. Prod. Lett.* **1995**, *6*, 295. For formal synthesis of sphingofungin D, see: (i) Mori, K.; Otaka, K. *Tetrahedron Lett.* **1994**, *35*, 9207.

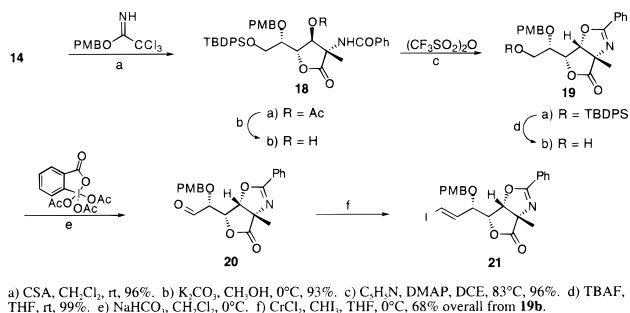
(9) (a) Cha, J. K.; Kim, N.-S. *Chem. Rev.* **1995**, *95*, 1761. (b) Cha, J. K.; Christ, W. J.; Kishi, Y. *Tetrahedron* **1984**, *40*, 2247.

(10) Trost, B. M.; Ariza, X. *Angew. Chem., Int. Ed. Engl.* **1997**, *36*, 2635.

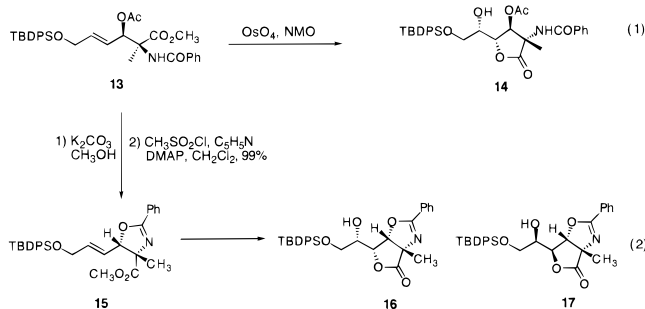
(11) Trost, B. M.; Lee, C. B.; Weiss, J. M. *J. Am. Chem. Soc.* **1995**, *117*, 7247.

(12) Quimper, M.; Ruest, L.; Deslongchamps, P. *Synthesis* **1992**, 132.

(13) Trost, B. M.; Van Vranken, D. L.; Bingle, C. J. *J. Am. Chem. Soc.* **1992**, *114*, 9327. For a review, see: Trost, B. M. *Acc. Chem. Res.* **1996**, *29*, 355.

Scheme 2. Asymmetric Synthesis of Polar Head Group from **14**

at $-5\text{ }^{\circ}\text{C}$ in THF gave a 10.5:1 ratio of **10** and **11**, isolated in 70% and 5% yields respectively. Both diastereomers have an ee of 89% as determined by chiral HPLC. Methanolysis (1% CSA or *p*-TsOH, CH₃OH, room temperature) of **10** gave a quantitative yield of the protected amino acid **13**. Dihydroxylation of alkene **13** (eq 1, 2% OsO₄, NMO)¹⁴ proceeded best in moist methylene chloride to give a single product **14** in which the initial diol spontaneously lactonized. In contrast to the excellent diastereoselectivity



selectivity in the dihydroxylation of **13** to give **14**, inverting the C-3 hydroxyl group first to form **15** and then performing the dihydroxylation (eq 2) gave a 1:5 ratio of **16** and **17** in 95% yield. Asymmetric dihydroxylation (AD-mix- α)¹⁵ was unable to overcome this latter substrate controlled diastereoselectivity (1:2.3, 100% yield).

The relative stereochemistry of **17** was established by X-ray crystallography and therefore allows the assignment of the minor lactone as shown in **16**. Correlation of **14** with **16** therefore establishes the relative configuration of **14**.¹⁶ The absolute configuration for asymmetric alkylations of **9** was established by the *O*-methyl mandelate method¹⁷ and, in this case, was ultimately verified by correlation of our synthetic product to the natural material.

Scheme 2 outlines the synthesis of the fully elaborated head fragment. Setting the proper configuration at C-3 was achieved

(14) VanRheenen, V.; Kelly, R. C.; Cha, D. Y. *Tetrahedron Lett.* **1976**, 1973. Also see ref 9.

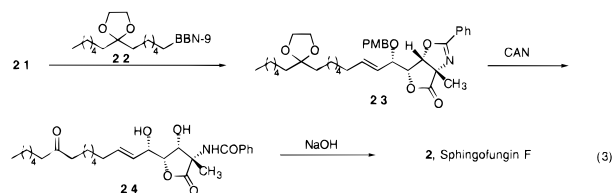
(15) Kolb, H. C.; Van Nieuwenhze, M. S.; Sharpless, K. B. *Chem. Rev.* **1994**, *94*, 2483. Kolb, H. C.; Anderson, P. G.; Sharpless, K. B. *J. Am. Chem. Soc.* **1994**, *116*, 1278. Sharpless, K. B.; Amberg, W.; Bennani, Y. L.; Crispino, G. A.; Hartung, J.; Jeong, K.-S.; Kwong, H.-L.; Morikawa, K.; Wang, Z.-M.; Xu, D.; Zhang, X.-L. *J. Org. Chem.* **1992**, *57*, 2768.

(16) **19a** was converted into **16** in 83% yield (DDQ, aqueous CH₂Cl₂, 0 $^{\circ}\text{C}$).

(17) Trost, B. M.; Belletire, J. L.; Godleski, S.; McDougal, P. G.; Balkovec, J. M.; Baldwin, J. J.; Christy, M. E.; Ponticello, G. S.; Varga, S. L.; Springer, J. P. *J. Org. Chem.* **1986**, *51*, 2370.

by inversion with neighboring group participation by simple activation of the hydroxyl group of **18b**, which directly formed oxazoline **19a**. The aldehyde **20** underwent the iodomethylenation with low valent chromium to generate the *E*-alkene **21** exclusively.¹⁸

The final stage of the synthesis (eq 3) added all of the remaining carbon atoms by Suzuki cross-coupling (5% (dppf)PdCl₂, 5% Ph₃As, Cs₂CO₃, DMF-THF-H₂O, room temperature)¹⁹ with the organoborane **22** formed *in situ* by hydroboration of the ethylene ketal of alkene **5** (derived in 3 steps from commercially available heptanoyl chloride)²⁰ to give alkene **23** in 94% yield. Oxidative



cleavage of the PMB group (CAN, CH₃CN, H₂O, room temperature, 93% yield) effected simultaneous hydrolyses of the ketal and the oxazoline to give keto alkene **24**. Base hydrolysis (1 N NaOH, reflux) and neutralization with Amberlite IRC-76 produced sphingofungin F, mp 143–5 $^{\circ}\text{C}$ (lit. mp 142–4 $^{\circ}\text{C}$), identical spectroscopically to the natural product. Our [α]_D of +0.99 (*c* 0.25, CH₃OH) compared to the reported [α]_D +0.8 (*c* 0.33, CH₃-OH) confirms the identity of the absolute configuration.

The efficiency of this synthesis is noted in that it requires only 15 linear steps from commercially available *cis*-2-butene-1,4-diol and proceeds in 17% overall yield. All the stereochemistry derives from that established in the asymmetric palladium catalyzed alkylation. It is noteworthy that the route provides ready access to a number of diastereomers. For example, the epimer at C-3 would derive by taking **14** through the synthesis without inversion. Access to **17** provides the epimers at C-4 and C-5. Variation of the azlactone would also allow variation of the alkyl substituent. Finally, the lipid tail can readily be varied in the cross-coupling reaction. The success of the palladium-catalyzed alkylation demonstrates a high level of catalyst control of diastereoselectivity, further demonstrating the potential selectivity that the complexes bearing these bidentate ligands like **12** may exert beyond enantioselectivity.

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Supporting Information Available: Experimental procedures and characterization data for **10**, **11**, **13–19**, **21**, **23**, **24**, and **2** (15 pages, print/PDF). See any current masterhead page for ordering information and Web access instructions.

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(18) Takai, T.; Nitta, K.; Utimoto, K. *J. Am. Chem. Soc.* **1986**, *108*, 7408. Kende, A. S.; DeVita, R. J. *Tetrahedron Lett.* **1990**, *31*, 307.

(19) Miyaura, N.; Suzuki, A. *Chem. Rev.* **1995**, *95*, 2457. Miyaura, N.; Ishiyama, T.; Sasaki, H.; Ishikawa, M.; Satoh, M.; Suzuki, A. *J. Am. Chem. Soc.* **1989**, *111*, 314. Ohba, M.; Kawase, N.; Fujii, T. *J. Am. Chem. Soc.* **1996**, *118*, 8250. Johnson, C. R.; Braun, M. P. *J. Am. Chem. Soc.* **1993**, *115*, 11014.

(20) Preparation of the ethylene ketal of alkene **5**:

